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TRANSMITTAL LETTER TO THE UNITED STATES DESIGNATED/ELECTED OFFICE (DO/EO/US) CONCERNING A FILING UNDER 35 U.S.C. 371		U.S. APPLICATION NO. (if known, see 37 CFR 1.5) 09/743364
INTERNATIONAL APPLICATION NO. PCT/NL99/00442	INTERNATIONAL FILING DATE 12.07.99 (July 12, 1999)	PRIORITY DATES CLAIMED 10.07.98 (July 10, 1998)
TITLE OF INVENTION CHEMOTAXIS-INHIBITING PROTEIN OF STAPHYLOCCUS (CHIPS) AND ITS USE		
APPLICANT(S) FOR DO/EO/US Johannes A. G. VAN STRIJP, Cornelis P. M. VAN KESSEL		
Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:		
<ol style="list-style-type: none"> 1. <input checked="" type="checkbox"/> This is a FIRST submission of items concerning a filing under 35 U.S.C. 371. 2. <input type="checkbox"/> This is a SECOND or SUBSEQUENT submission of items concerning a filing under 35 U.S.C. 371. 3. <input checked="" type="checkbox"/> This express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1) 4. <input checked="" type="checkbox"/> A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date 5. <input checked="" type="checkbox"/> A copy of the International Application as filed (35 U.S.C. 371(c)(2)) <ol style="list-style-type: none"> a. <input type="checkbox"/> is transmitted herewith (required only if not transmitted by the International Bureau) b. <input checked="" type="checkbox"/> has been transmitted by the International Bureau. c. <input type="checkbox"/> is not required, as the application was filed in the United States Receiving Office (RO/US). 6. <input type="checkbox"/> A translation of the International Application into English (35 U.S.C. 371(c)(2)). 7. <input checked="" type="checkbox"/> Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3)) <ol style="list-style-type: none"> a. <input type="checkbox"/> are transmitted herewith (required only if not transmitted by the International Bureau). b. <input type="checkbox"/> have been transmitted by the International Bureau c. <input type="checkbox"/> have not been made, however, the time limit for making such amendments has NOT expired d. <input checked="" type="checkbox"/> have not been made and will not be made 8. <input type="checkbox"/> A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)) 9. <input type="checkbox"/> An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)) 10. <input type="checkbox"/> A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)) 		
Items 11. to 16. below concern document(s) or information included:		
<ol style="list-style-type: none"> 11. <input type="checkbox"/> An Information Disclosure Statement under 37 CFR 1.97 and 1.98 12. <input type="checkbox"/> An assignment document for recording A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included 13. <input checked="" type="checkbox"/> A FIRST preliminary amendment <input type="checkbox"/> A SECOND or SUBSEQUENT preliminary amendment. 14. <input type="checkbox"/> A substitute specification 15. <input type="checkbox"/> A change of power of attorney and/or address letter 16. <input checked="" type="checkbox"/> Other items or information <ol style="list-style-type: none"> a. WO 00/02913-Front Page with Abstract, specification, claims and drawings (24 pp.) b. Search Report (3 pp.) c. International Preliminary Examination Report (9 pp.) 		

U.S. APPLICATION NO. (if known) 09/743364 <small>(37 CFR 1.53)</small>		INTERNATIONAL APPLICATION NO. PCT/NL99/00442		ATTORNEY'S DOCKET NUMBER 702-002197	
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17. <input checked="" type="checkbox"/> The following fees are submitted: BASIC NATIONAL FEE (37 CFR 1.492(a)(1)-(5)): Search Report has been prepared by the EPO or JPO \$860.00 International preliminary examination fee paid to USPTO (37 CFR 1.482) \$690.00 No international preliminary examination fee paid to USPTO (37 CFR 1.482) but international search fee paid to USPTO (37 CFR 1.445(a)(2)) \$710.00 Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$1000.00 International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(2)-(4) \$100.00				CALCULATIONS PTO USE ONLY	
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Surcharge of \$130.00 for furnishing the oath or declaration later than <input type="checkbox"/> 20 <input checked="" type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(e)).				\$ 130.00	

CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE		
Total claims	15 - 20	0	X \$18.00	\$ 0.00	
Independent claims	11 - 3 =	8	X \$80.00	\$ 640.00	
MULTIPLE DEPENDENT CLAIM(S) (if applicable)			+ \$270.00	\$ 0.00	
TOTAL OF ABOVE CALCULATIONS =				\$ 1630.00	
Reduction of 1/2 for filing by small entity, if applicable. Small Entity Statement verified by Applicant(s) attorney.				\$ 0.00	
SUBTOTAL =				\$ 1630.00	
Processing fee of \$130.00 for furnishing the English translation later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(f)).				\$ 0.00	
TOTAL NATIONAL FEE =				\$ 1630.00	
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property				\$ 0.00	
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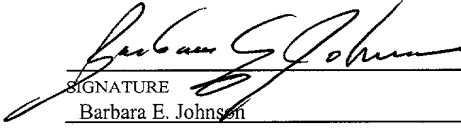
a. ☒ A check in the amount of **\$ 1630.00** to cover the above fees is enclosed

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c. ☒ The Assistant Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. 23-0650. A duplicate copy of this sheet is enclosed.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

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09/743364
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PATENT APPLICATION/PCT
Attorney Docket No. 702-002197

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of :

Johannes A. G. VAN STRIJP, : **CHEMOTAXIS-INHIBITING**
Cornelis P. M. VAN KESSEL : **PROTEIN OF *STAPHYLOCCUS***
 : **(CHIPS) AND ITS USE**

International Application :
No. PCT/NL99/00442 :

International Filing Date :
12 July 1999 :

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10 July 1998 :

Serial No. Not Yet Assigned :

Filed Concurrently Herewith :

Pittsburgh, Pennsylvania
January 9, 2001

PRELIMINARY AMENDMENT

BOX PCT

Assistant Commissioner for Patents
Washington, DC 20231

Sir:

Prior to initial examination, please amend the above-identified patent application
as follows:

Page 1, after the title, insert the following headings:

--BACKGROUND OF THE INVENTION

1. Field of the Invention--.

Page 1, after line 6, insert the following heading:

--2. Description of the Prior Art--.

Page 3, before line 1, insert the following heading:

--SUMMARY OF THE INVENTION--.

Page 8, after line 6, insert the following heading:

--BRIEF DESCRIPTION OF THE DRAWINGS--

Page 9, delete line 1 in its entirety and substitute the following heading therefor:

--DESCRIPTION OF THE PREFERRED EMBODIMENTS--.

IN THE CLAIMS:

Original claims 1-14 were amended during Chapter II proceedings by substituting new claims 1-16 in a letter dated August 28, 2000. Please cancel original claims 1-14 and cancel amended claims 1-16, and add new claims 17-31 as follows:

--17. A chemotaxis-inhibiting protein of *Staphylococcus* (CHIPS protein), which is characterized by:

- a) a molecular weight of about 17 kD;
- b) the N-terminal amino acid sequence as given in figure 4 (SEQ ID NO: 1);

5 and

c) a biological activity which consists of the capacity to prevent the binding of fMLP and/or C5a to granulocytes in a test as described in example 1, under 1.2, and fragments thereof that have the biological activity as defined under c).

18. A biologically active substance comprising a substance selected from the group consisting of the CHIPS protein and biologically active fragments thereof.

19. A medicine comprising a substance selected from the group consisting of the CHIPS protein and biologically active fragments thereof.

20. A method of treatment of acute and chronic inflammation reactions and HIV infection comprising administration of a substance selected from the group consisting of the CHIPS protein and biologically active fragments thereof.

21. Antibodies against a substance selected from the group consisting of the CHIPS protein and biologically active fragments thereof.

22. A method for the treatment of *Staphylococcus* infection comprising the administration of antibodies against a substance selected from the group consisting of the CHIPS protein and biologically active fragments thereof.

23. A therapeutic composition comprising a suitable excipient and a substance selected from the group consisting of the CHIPS protein and biologically active fragments thereof.

24. The composition as claimed in claim 23 for treating acute and chronic inflammation reactions and HIV infection.

25. A therapeutic composition comprising a suitable excipient and one or more antibodies against a substance selected from the group consisting of the CHIPS protein and biologically active fragments thereof.

26. A method of purifying the CHIPS protein as claimed in claim 17, comprising the steps of:

a) guiding over an absorption chromatography column the culture supernatant of *Staphylococcus aureus* or a liquid obtained therefrom after pre-purification;

b1) subsequently guiding the flow-through of the absorption chromatography column first over an affinity chromatography column and thereafter guiding the eluate of the affinity chromatography column over a DNA column; or

b2) subsequently guiding the flow-through of the absorption chromatography column first over a DNA column and thereafter guiding the flow-through of the DNA column over an absorption chromatography column; and

c) guiding the flow-through of the last column of step b1) respectively the eluate of the last column of step b2) over a gel filtration column and selecting the fraction with a molecular weight of about 17 kD.

27. The method as claimed in claim 26, wherein the affinity chromatography column is a Ligand Dye "yellow" column, the absorption chromatography column is a Ligand Dye "green" column and the DNA column a DNA cellulose column.

28. A method of determining the activity of the CHIPS protein and/or the biologically active fragments thereof as claimed in claim 17 or proteins with an analogous function, comprising the steps of:

- a) introducing into a first compartment labeled cells capable of chemotaxis,
5 in particular leucocytes;
- b) introducing one or more chemoattractants into a second compartment separated from the first compartment by a membrane permeable to at least the cells;
- c) placing the protein for testing into the first compartment; and
- d) measuring the quantity of label in the second compartment after a
10 determined time.

29. The method of determining the chemotaxis-modulating activity of a substance, comprising the method steps as claimed in claim 28, wherein the substance for testing replaces the protein of step c).

30. A method of determining the activity of the CHIPS protein and/or the biologically active fragments thereof as claimed in claim 17 or molecules, such as proteins, with an analogous activity, comprising the steps of:

- a) incubating granulocytes suspended in a medium with CHIPS-containing
5 material for a determined time;
- b) washing the granulocytes with fresh medium and resuspending the granulocytes in such medium;

- 10 c) incubating the granulocytes with fMLP and/or C5a that is labeled with a detectable label in order to effect binding of the labeled fMLP and/or C5a to the granulocytes;
- d) washing away the unbound detectable label; and
- e) analysing the binding of the labeled fMPL and/or C5a to the granulocytes by measuring the detectable label.

31. The method as claimed in claim 30, comprising the steps of:

- a) incubating granulocytes suspended in RPMI medium with 0.05% Human Serum Albumin (RPMI/HSA) with CHIPS-containing material for 30 min. at 37°C;
- b) placing the granulocytes on ice and washing them once in RPMI/HSA at 4°C;
- c) resuspending the granulocytes in fresh RPMI/HSA medium;
- d) incubating the granulocytes with fluorescently labeled fMLP and/or C5a in order to effect binding of the labeled fMLP and/or C5a to the granulocytes;
- e) washing away the unbound fluorescent label; and
- f) analysing the binding of the fluorescent fMPL and/or C5a to the granulocytes by measuring the fluorescence.--

IN THE ABSTRACT:

After the claims, please insert a page containing the Abstract Of The Disclosure, which is attached hereto as a separately typed page.

REMARKS

The specification has been amended by this Preliminary Amendment to place the application in conformance with standard United States patent practice.

Under date of August 28, 2000, Applicants submitted substitute sheets containing amended claims 1-16 for the above-identified PCT application. Original claims 1-14 have been canceled by this Preliminary Amendment and amended claims 1-16 have been cancelled and rewritten as new claims 17-31 to eliminate the multiple dependencies and to bring the claims into conformance with standard United States Patent practice.

A further Amendment will be submitted in due course to address the amino acid sequence listings.

An Abstract of the Disclosure has been added as a separately typed page to be inserted after the claims.

Entry of this Preliminary Amendment and examination and allowance of claims 17-31 are respectfully requested.

Respectfully submitted,

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CHEMOTAXIS-INHIBITING PROTEIN OF STAPHYLOCOCCUS (CHIPS)
AND ITS USE

The present invention relates to a new protein which can be used in the treatment of inflammation reactions. The invention further relates to a method of purifying the protein. Finally, the invention relates to
5 a screening test with which analogous proteins can be detected.

An inflammation reaction is very generally a process wherein defence cells make their way to a source of infection and there ensure the elimination of the
10 cause. Different mediators are herein released which contribute to elimination but also produce the inflammation symptoms. A distinction can be made between acute inflammations (such as sepsis) and latent chronic inflammations (such as rheumatism). In people with a
15 lowered resistance acute inflammations can occur more often and more severely (as in the case of AIDS).

An infection with bacteria results in the formation of chemotactic factors which ensure that the leucocytes go to the source of the infection. A
20 chemotactic substance is present in a gradient along which the leucocytes move in a directed manner. The source of a chemotactic agent can be the bacteria itself. These agents are for instance small proteins with a terminal formyl group, such as fMLP (N-formyl-Methionyl-Leucyl-Phenylalanine). Other chemotactic agents are
25 activated complement factors (the anaphyloxins C3a and C5a), leukotrienes (such as LTB4 (Leukotriene B4) and PAF (Platelet-Activating Factor) and chemokines produced by different cell types such as interleukin-8 (monocytes and
30 endothelial cells), RANTES (Regulated upon Activation, Normal T-cell Expressed and Secreted), eotaxin, MCP (Monocyte Chemotactic Protein), MIP (Macrophage Inflammatory Protein) and others.

Some chemokines are only specific to a
35 determined type of leucocyte, others affect a plurality

of cells. The receptors for chemokines are subdivided into two main groups, the CC and CXC receptors which all belong to the serpentine, receptors which traverse the membrane 7 times. The serpentine are rhodopsin-like GTP-binding protein linked receptors.

The super family of chemotactic cytokines, chemokines, is characterized by 4 conserved cysteines. Depending on the relative position of the first two cysteines, two families can be distinguished: the CXC or alpha-chemokines and the CC or beta-chemokines. The CXC chemokines are particularly active on granulocytes while the CC chemokines activate a wide range of leucocytes including monocytes, eosinophils, T-lymphocytes, NK cells and dendritic cells. This family of chemokines and also the classical chemotactic agents such as fMLP and C5a bind and activate the serpentine.

Neutralization of chemokines has already been applied experimentally, in particular the administering of antibodies against IL-8 was found to be effective in a number of animal experimental inflammation models. In addition, it has recently been demonstrated that a number of chemokine receptors (CCR5 and CXCR4 in particular) play a part as co-factor in the infection of cells by HIV. For the T-cell-trophic strains of HIV, CXCR4 has been identified as co-factor and for monotrophic strains this is CCR5. Blocking of these receptors with antibodies or ligands inhibited the HIV infection in in vitro models. Moreover, people with a genetic variant of the CCR5 receptor consisting of a 32 base pair deletion are found to be resistant to infection with HIV.

In addition to induction of chemotaxis, the directed migration of the leucocytes, in low concentrations a number of chemokines are also potent activators or primers of other leucocyte functions. It is therefore desirable to achieve a blocking of chemokines whereby inflammation reactions can be kept in check.

It is therefore the object of the present invention to provide a new agent with inflammation-inhibiting properties for the treatment of acute and chronic inflammation reactions and HIV.

5 During the research which led to the present invention a protein was found in the extracellular medium of growing Staphylococcus aureus (S. aureus) which was found capable of blocking different chemokine receptors. Incubation of different cells with the medium resulted in
10 a greatly reduced expression of a number of the chemokine receptors, both of the expression of receptors of classical chemotactic agents such as fMLP and C5a on granulocytes and of the expression of CXCR4 and CCR5 receptors on lymphocytes, monocytes and macrophages. The
15 reduced receptor expression was related to greatly reduced chemotaxis relative to the chemokines, as well as a reduced infection with HIV.

 The activity of the protein is already manifest in the culture supernatant of the growing S.aureus.
20 According to the invention however, the active protein was also purified by means of a number of Ligand Dye columns. A pre-purification was first performed on a so-called "yellow column" ("Reactive Yellow 86" ligand dye cross-linked 4% beaded agarose column (Sigma)), followed
25 by an absorption chromatography column (the so-called "green column" ("Reactive Green 19" ligand dye cross-linked 4% beaded agarose column (Sigma)) and a DNA column (DNA Cellulose (Pharmacia)). Both latter columns can be interchanged. The DNA column removes a contaminant with
30 the same molecular weight as the protein according to the invention. The absorption chromatography column concentrates the protein and is selective for the protein. Finally, a post-purification also takes place by means of gel filtration and optionally a concentration.
35 In the gel filtration the protein with the molecular weight of about 17 kD is selected. This is the protein according to the invention.

Each step in the purification method can be monitored by means of testing the activity of the flow-through or the eluate of the different columns. This takes place by monitoring whether the flow-through or the eluate is able to prevent the binding of fMLP to granulocytes. An extensive test protocol is given in the examples.

The first (N-terminal) 15 amino acids of the purified protein were determined by means of micro-sequencing. This sequence is given in figure 4. With sequence analysis no homology with any known bacterial or eucaryotic amino acid sequence was found in databases. This is therefore a new and unique protein.

Because this protein is isolated from the supernatant of the Staphylococcus aureus and gives inhibition of chemotaxis, this protein is herein also designated as "CHIPS": CHEmotaxis Inhibitory Protein from Staphylococcus aureus.

The present invention therefore relates according to a first aspect thereof to the CHIPS protein, which is characterized by:

- a) a molecular weight of about 17 kD;
- b) the N-terminal amino acid sequence as given in figure 4; and
- c) a biological activity which consists of the capacity to prevent the binding of fMLP to granulocytes in a test as described in example 1, and biologically active fragments thereof.

The CHIPS protein influences the chemotaxis of leucocytes to the source of the chemoattractant, such as the bacteria. It has been found according to the invention that the number of at least two receptors (fMLP and C5a) on the leucocytes, in particular granulocytes, is down-regulated. The down-regulation is reversible.

The invention further relates to a therapeutic composition, comprising a suitable excipient and the CHIPS protein and/or biologically active fragments

thereof. The composition can be used for the treatment of acute and chronic inflammation reactions and HIV infection. The protein ensures that the receptors which provide movement of the leucocyte to the chemotactic substance are blocked.

The invention likewise relates to the CHIPS protein and/or biologically active fragments thereof for use in the treatment of acute and chronic inflammation reactions and HIV infection, as well as the use of the CHIPS protein and/or biologically active fragments thereof for the manufacture of a therapeutic preparation for the treatment of said symptoms.

According to a subsequent aspect of the invention antibodies against the CHIPS protein and/or fragments thereof are provided for use in the treatment of staphylococcus infection. The proteins will block the activity of CHIPS or its fragments and thus restart the chemotaxis terminated by the bacteria, whereby the natural defence against the bacteria is restored.

The therapeutic compositions, which according to the invention contain CHIPS or antibodies thereagainst as active ingredient, will be particularly intended for parenteral, and then specifically, intravenous use. The therapeutic compositions can be prepared by combining (i.e. mixing, dissolving etc.) CHIPS with pharmaceutically acceptable excipients suitable for intravenous administration. The concentration of the active ingredient in a therapeutic composition can vary between 0.001% and 100%, depending on the nature of the treatment and the method of administration. The dose of the active ingredient for administering likewise depends on the administering route and application, but may for instance vary between 0.001 and 1 mg per kg of body weight, preferably between 1 μ g and 100 μ g per kg of body weight.

The invention further relates to a method of purifying the CHIPS protein, comprising of:

a) guiding over an absorption chromatography column the culture supernatant of Staphylococcus aureus or a liquid obtained therefrom after pre-purification;

b1) subsequently guiding the flow-through of the absorption chromatography column first over an affinity chromatography column and thereafter guiding the eluate of the affinity chromatography column over a DNA column; or

b2) subsequently guiding the flow-through of the absorption chromatography column first over a DNA column and thereafter guiding the flow-through of the DNA column over an absorption chromatography column;

c) guiding the flow-through respectively the eluate of the last column of step b) over a gel filtration column and selecting the fraction with a molecular weight of about 17 kD.

"Flow-through" is herein understood to mean that part of the loaded liquid having situated therein the constituents which come from the column without extra treatment. The constituents in this flow-through do not bind to the column. "Eluate" is understood to mean the liquid which comes from the column after elution and which contains the constituents from the liquid loaded on the column which were bound to the column and were released again therefrom by the elution. In the method according to the invention the absorption column binds most constituents of the loaded culture medium or a liquid obtained therefrom after pre-purification. The affinity column binds CHIPS and the Snase (Staphylococcal Nuclease) which has the same molecular weight as CHIPS and the same affinity (or lack thereof) for the affinity column respectively the absorption column. The DNA column binds only the Snase, whereby this is separated from CHIPS.

This method works particularly well if the first affinity chromatography column is a so-called Ligand Dye "yellow" column, the second affinity

chromatography column is a so-called Ligand Dye "green" column and the DNA column a DNA cellulose column.

Finally, the invention also relates to a determination or assay for determining the activity of the CHIPS protein or proteins with an analogous function, comprising of:

a) introducing into a first compartment labelled cells capable of chemotaxis, in particular leucocytes,

b) introducing one or more chemoattractants into a second compartment separated from the first compartment by a membrane permeable to at least the cells,

c) placing the protein for testing into the first compartment;

d) measuring the quantity of label in the second compartment after a determined time.

The cells are capable of moving through the membrane in the direction of the chemoattractant. The presence of CHIPS or an analogous protein prevents the migration by deactivating the receptor(s) for the chemoattractant(s). This test can be applied more generally to also determine the chemotaxis-modulating activity of other substances. The method steps are then the same.

Proteins analogous to CHIPS which are found in this manner can be subjected to the same purification as CHIPS to thus be able to determine homology between the two.

The CHIPS protein according to the invention has also been found in S.epidermis as well as in S.aureus.

The manner in which CHIPS was isolated from the culture supernatant of Staphylococcus aureus via the steps of ligand dye affinity and absorption chromatography, gel filtration and concentration, in addition to the testing of the activity of CHIPS on

different chemokine receptors on different leucocytes and the testing of the inhibition of HIV infection in T-cells and macrophages is described in the examples following hereinbelow, which are only intended by way of

5 illustration and are not intended to limit the invention in any way whatsoever.

Reference is made in the examples to the following figures:

Figure 1 shows the incubation of granulocytes
10 with a concentration series of a Staphylococcus supernatant (SaS) and the effect on the fMLP receptors and the C5a receptors.

Figure 2 shows the effect of a concentration series of SaS on the chemotaxis of granulocytes to fMLP.

15 **Figure 3** shows the fractions of the gel filtration column with the optical density (OD) (line with diamonds) and the activity in the fMLP receptor assay as a percentage of inhibition (bars).

Figure 4 shows the sequence of the first 15 (N-
20 terminal) amino acids of CHIPS (of the estimated 125 in total).

Figure 5a shows the incubation of granulocytes with a concentration series of purified CHIPS and the effect on the fMLP receptors and the C5a receptors.

25 **Figure 5b** shows the incubation of granulocytes with a concentration series of purified CHIPS and the effect on the directed migration to fMLP.

Figure 6 shows the expression of CXCR4 on lymphocytes.

EXAMPLES

EXAMPLE 1

Identification and isolation of CHIPS as chemotaxis- 5 inhibiting molecule

MATERIAL AND METHOD

1.1 Isolation of the protein

Staphylococcus aureus 1690 (a clinical isolate, Utrecht Teaching Hospital) or Staphylococcus aureus Newman (van Dr TJ Foster, Dublin) is cultured overnight in IMDM medium (Gibco) and subsequently diluted 1:40 in fresh IMDM for a 7 hour culture at 37°C. After pelleting of the bacteria the S.aureus supernatant (referred to as SaS) is collected, filtered over a 0.2 µm filter and immediately used further. See also Veldkamp et al, Inflammation 21, 541-551 (1997).

A quantity of 2 litres of SaS is guided over three columns (25 ml) coupled in tandem. These three columns are successively a "Reactive Yellow 86" ligand dye cross-linked 4% beaded agarose column (Sigma), a DNA Cellulose (Pharmacia) and a "Reactive Green 19" ligand dye cross-linked 4% beaded agarose column (Sigma). After washing the green (Reactive Green 19) column is eluted with 2 M NaCl and the active fractions are pooled and concentrated 10x with polyethylene glycol. The concentrated material is separated on a Pharmacia Superdex-200 gel filtration column, whereafter the active fractions are pooled, concentrated, dialysed and freeze-dried. The final purified material is resuspended in a small volume of sterile water and used inter alia for micro-sequence analysis.

For this purpose a sample is analysed on a 12.5% SDS-PAGE (Mini-Protean II; BioRad) and transferred to Immobilon-P PVDF membrane (Millipore) by means of the Mini Trans-Blotter (BioRad). The proteins are stained with Coomassie Blue and the protein around 17 kD is excised. The N-terminal amino acid sequence of this

sample is determined in accordance with the automated Edman procedure, wherein use is made of a Perkin Elmer/Applied Biosystems 476A. Amino acid derivatives are analysed by means of HPLC.

5

1.2 Binding of fMLP to granulocytes

Granulocytes are isolated from heparinized blood of healthy volunteers via a Histopaque-Ficoll gradient in accordance with the standard method

10 (Troelstra et al, J. Leukocyte Biol. 61:173-178 (1997).

The remaining erythrocytes in the granulocyte fraction are lysed with sterile water (for 30 sec) and washed after recovery of the isotonicity. The cells are finally resuspended in RPMI (Gibco) with 0.05% Human Serum

15 Albumin (RPMI/HSA).

In Falcon tubes 50 μ l cells (5×10^6 cells/ml) are incubated with 50 μ l CHIPS-containing material (SaS, purified CHIPS or column fractions) for 30 min at 37°C.

The cells are placed on ice and washed once RPMI/HSA (at
20 4°C) and resuspended in 50 μ l fresh medium. 5 μ l BODIPY-labelled fMLP (end concentration 0.1 μ M; Molecular Probes) is then added and the sample is incubated for 60 minutes on ice. After washing the fluorescent fMLP binding to the granulocytes is analysed with a

25 flowcytometer (FACScan; Becton Dickinson). The average fluorescence value of 5000 granulocytes is calculated with Lysis II software.

1.3 Chemotaxis

30 In order to determine the directed migration use is made of a Transwel system (Costar) consisting of an upper compartment and a lower compartment separated by a 3 μ m polycarbonate membrane. The granulocytes are labelled with BCECF (2-carboxyethyl-5-(and-6-)
35 carboxyfluorescein; Molecular Probes), a fluorescent label which enters the cytoplasm of the cells. The cells (5×10^6) are incubated for 20 minutes at 22°C with 3 μ M

BCECF-AM (the acetomethyl ester of 2-carboxyethyl-5-(and-6-)-carboxyfluorescein), subsequently washed three times and resuspended to 5×10^6 cells/ml in RPMI/HSA. 100 μ l of cells and the desired quantity of the CHIPS protein is introduced into the upper compartment of the Transwell system and the whole is suspended in the wells of a standard 24-well microtitre plate (Costar). Each well contains 600 μ l RPMI/HSA with or without addition of the chemoattractant for testing. The chemoattractants are: recombinant C5a (Sigma), recombinant interleukin-8 (Pepro Tech), Platelet Activating Factor-16 (PAF-16; Calbiochem) or fMLP (Sigma). After 60 minutes incubation at 37°C the Transwell container is lifted from the wells and the microtitre plate is analysed for fluorescence in a CytoFluorII (PerSeptiveBiosystems). The degree of fluorescence is a direct measure for the number of granulocytes which has migrated through the membrane and is expressed as a percentage of the fluorescence of the added number of cells.

RESULTS

Figure 1 shows the effect of the incubation of granulocytes with a concentration series of a Staphylococcus-supernatant (SaS) on the fMLP receptors and the C5a receptors. A strong down regulation of both the C5a and the fMLP receptor is visible.

Figure 2 shows that the chemotaxis (cell movement) to the attractant (fMLP) is strongly inhibited.

Figure 3 shows that the strongest inhibiting activity is situated in the elution range between 240 and 280 ml. The volume fractions correspond here with a protein of about 17 kD.

Figure 4 shows the sequence of the first 35 (N-terminal) amino acids of CHIPS (of the estimated 125 in total). On the basis of this sequence a synthetic peptide was made of the first 15 amino acids in accordance with standard Fmoc chemistry as described inter alia in De

Haas et al, J.Immunol. 161:3607-3615 (1998) and Alonso de Velasco et al, Infect. Immun. 62:799-808 (1994).

Antibodies generated against this peptide in rabbits (coupled to KLH in accordance with the instructions of the manufacturer, Pierce, and subcutaneously immunized with Freund's Complete Adjuvant, followed by a booster injection with Freund's Incomplete Adjuvant), as for instance described in Alonso de Velasco et al, supra, neutralize the activity of CHIPS.

10

EXAMPLE 2

Reduced expression of chemokine receptors on granulocytes due to CHIPS.

15 MATERIAL AND METHOD

2.1 Receptor expression

The expression of the different chemokine receptors is determined with specific fluorescent-labelled antibodies and flowcytometry. The procedure followed is analogous to that described under 1.2 in example 1. Use is made of the following monoclonals: S5/1, anti-CD88 (C5a receptor) from Serotec Ltd; SE2, anti-CDw128A (IL-8 receptor) from Alexis Corporation; anti-PAF Receptor from Alexis Corporation. After 25 incubation with CHIPS the cells are incubated for 30 min on ice with 5 µg/ml antibody and after washing are labelled with a F(ab)2-FITC-labelled goat anti-murine Ig (Dako).

The average fluorescence of the granulocytes is 30 a measure for the quantity of receptor on the cell surface. The relative expression, after subtraction of the background value, is expressed as a percentage of the cells which are incubated with control buffer.

RESULTS

Figure 5a shows that both the C5a receptor and the fMLP receptor also disappear from the surface of the cells due to purified CHIPS.

- 5 Figure 5b shows the incubation of granulocytes with a concentration series of purified CHIPS and the effect on the directed migration to fMLP. It can be seen that the chemotaxis (cell movement) to fMLP is inhibited completely and dose-dependently by purified CHIPS.

10

EXAMPLE 3

Reduced expression of chemokine receptors on T-cells due to CHIPS

15 MATERIAL AND METHOD

3.1 Receptor expression

- The mononuclear leucocytes (20% monocytes and 80% lymphocytes) are isolated from heparinized blood of healthy volunteers via a Ficoll gradient (Pharmacia) in accordance with the standard method (Antal-Szalmás et al, 20 J. Leukocyte Biol. 61, 721-728 (1997). After washing the cells are resuspended in RPMI/HSA. The procedure followed for measuring expression of the different chemokine receptors is the same as described under 2.1 in example 25 2. To measure the expression of the lymphotrophic co-receptor for HIV (CXCR4) use is made of monoclonal 12G5 anti-CXCR4 from Becton Dickinson.

RESULTS

- 30 Figure 6 shows that after incubation of mononuclear cells with CHIPS, the expression of CXCR4 on lymphocytes disappears.

CLAIMS

1. Chemotaxis-inhibiting protein of Staphylococcus (CHIPS protein), which is characterized by:

- a) a molecular weight of about 17 kD;
- 5 b) the N-terminal amino acid sequence as given in figure 4; and
- c) a biological activity which consists of the capacity to prevent the binding of fMLP to granulocytes in a test as described in example 1, and biologically
- 10 active fragments thereof.
2. CHIPS protein and/or biologically active fragments thereof as claimed in claim 1 for use as biologically active substance.
3. CHIPS protein and/or biologically active
- 15 fragments thereof as claimed in claim 1 or 2 for use as medicine.
4. CHIPS protein and/or biologically active fragments thereof as claimed in claim 1 for use in the treatment of acute and chronic inflammation reactions and
- 20 HIV infection.
5. Use of the CHIPS protein and/or biologically active fragments thereof as claimed in claim 1 for the manufacture of a therapeutic preparation for the treatment of acute and chronic inflammation reactions and
- 25 HIV infection.
6. Antibodies against the CHIPS protein and/or fragments thereof.
7. Antibodies as claimed in claim 6 for use in the treatment of staphylococcus infection.
- 30 8. Therapeutic composition comprising a suitable excipient and the CHIPS protein and/or biologically active fragments thereof.
9. Composition as claimed in claim 8 for treating acute and chronic inflammation reactions and HIV
- 35 infection.

10. Therapeutic composition comprising a suitable excipient and one or more antibodies against the CHIPS protein and/or biologically active fragments thereof.

5 11. Method of purifying the CHIPS protein, comprising of:

a) guiding over an absorption chromatography column the culture supernatant of Staphylococcus aureus or a liquid obtained therefrom after pre-purification;

10 b1) subsequently guiding the flow-through of the absorption chromatography column first over an affinity chromatography column and thereafter guiding the eluate of the affinity chromatography column over a DNA column; or

15 b2) subsequently guiding the flow-through of the absorption chromatography column first over a DNA column and thereafter guiding the flow-through of the DNA column over an absorption chromatography column;

c) guiding the flow-through of the last column of step b1) respectively the eluate of the last column of step b2) over a gel filtration column and selecting the fraction with a molecular weight of about 17 kD.

20 12. Method as claimed in claim 11, characterized in that the affinity chromatography column is a so-called Ligand Dye "yellow" column, the absorption chromatography column is a so-called Ligand Dye "green" column and the DNA column a DNA cellulose column.

30 13. Method of determining the activity of the CHIPS protein or proteins with an analogous function, comprising of:

a) introducing into a first compartment labelled cells capable of chemotaxis, in particular leucocytes,

35 b) introducing one or more chemoattractants into a second compartment separated from the first compartment by a membrane permeable to at least the cells,

c) placing the protein for testing into the first compartment;

d) measuring the quantity of label in the second compartment after a determined time.

- 5 14. Method of determining the chemotaxis-modulating activity of a substance, comprising the method steps as claimed in claim 13, wherein the substance for testing replaces the protein of step c).

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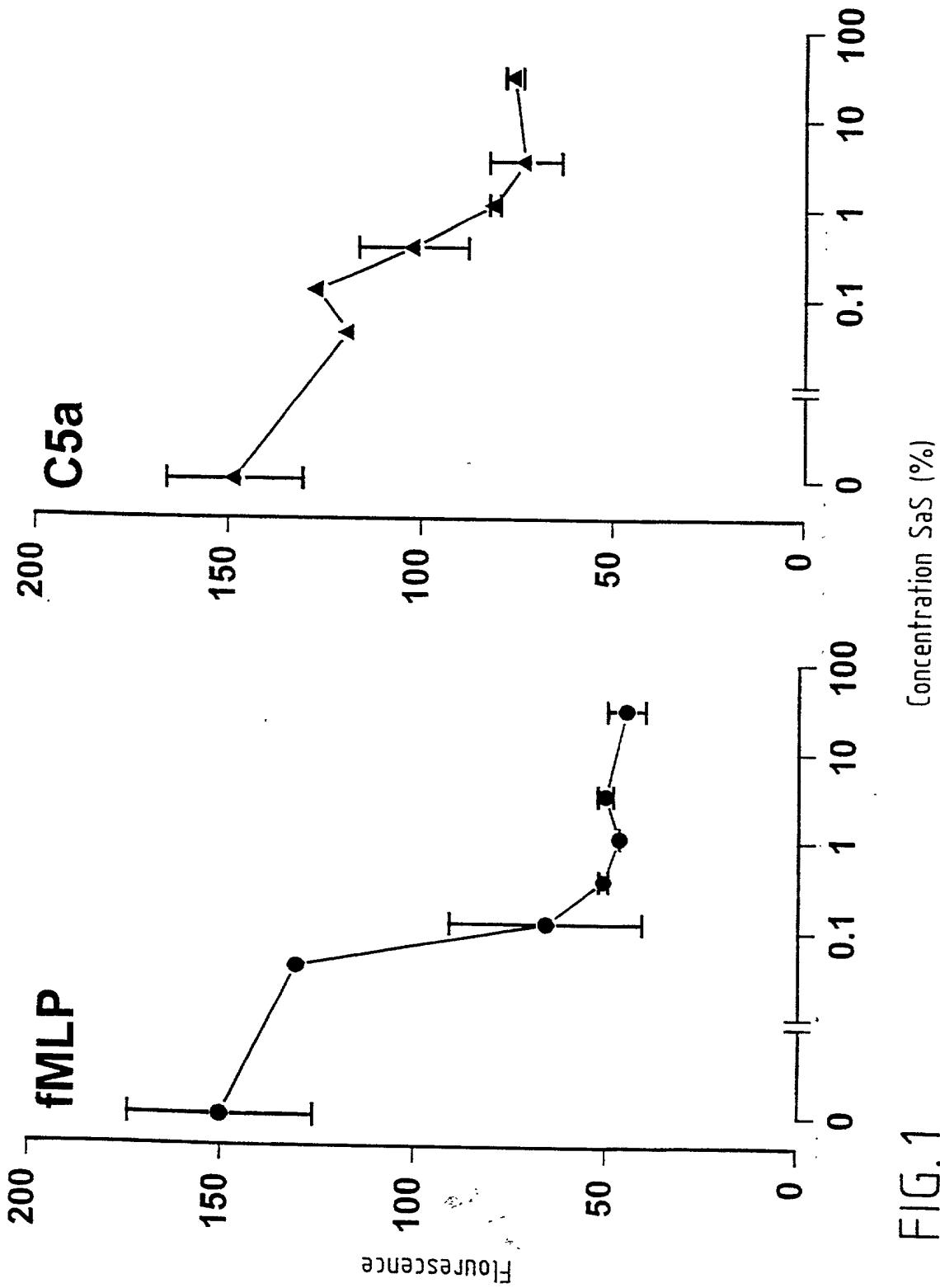


FIG. 1

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FIG. 2

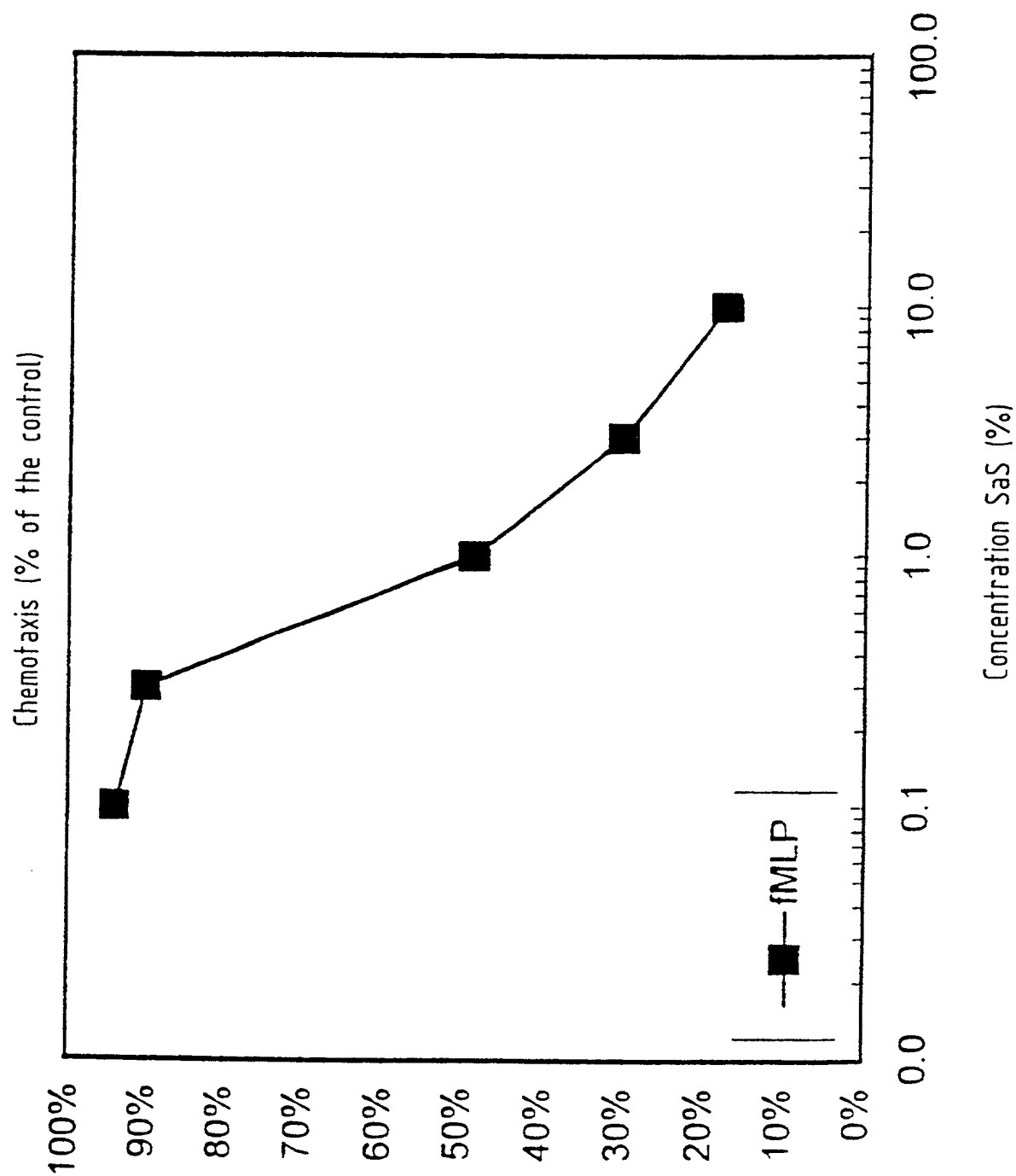
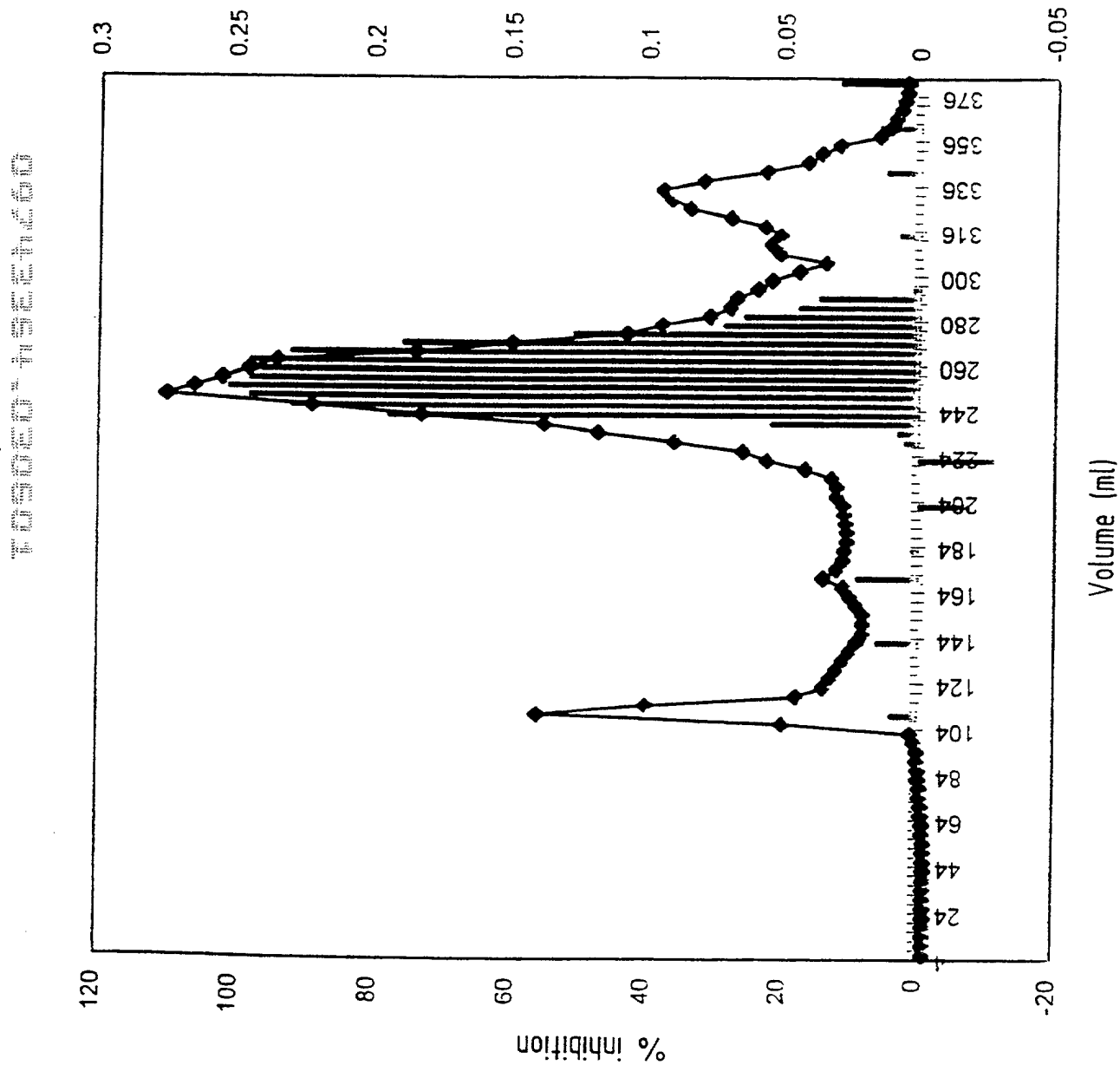


FIG. 3



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No	Letter	3-letter	Amino-acid
1	F	Phe	Phenylalanine
2	T	Thr	Threonine
3	F	Phe	Phenylalanine
4	E	Glu	Glutamate
5	P	Pr	Proline
6	F	Phe	Phenylalanine
7	P	Pro	Proline
8	T	Thr	Threonine
9	N	Asn	Asparagine
10	E	Glu	Glutamate
11	E	Glu	Glutamate
12	I	Ile	Isoleucine
13	E	Glu	Glutamate
14	S	Ser	Serine
15	N	Asn	Asparagine
16	K	Lys	Lysine
17	K	Lys	Lysine

18	M	Met	Methionine
19	L	Leu	Leucine
20	E	Glu	Glutamate
21	K	Lys	Lysine
22	E	Glu	Glutamate
23	K	Lys	Lysine
24	A	Ala	Alanine
25	Y	Tyr	Tyrosine
26	K	Lys	Lysine
27	E	Glu	Glutamate
28	S	Ser	Serine
29	F	Phe	Phenylalanine
30	K	Lys	Lysine
31	N	Asn	Asparagine
32	N?	Asn?	Asparagine?
33	G	Gly	Glycine
34	L	Leu	Leucine
35	P	Pro	Proline

FIG. 4

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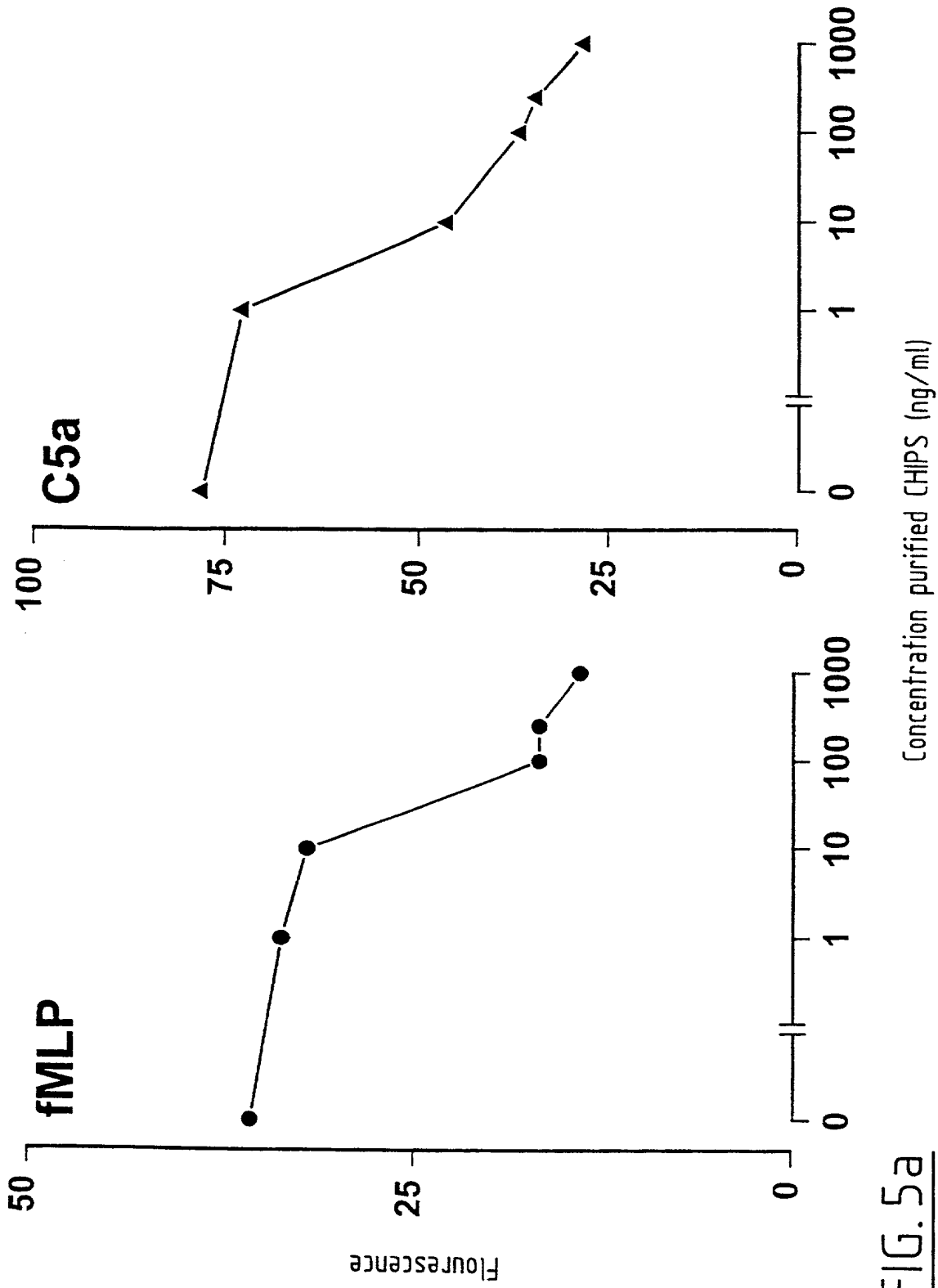


FIG. 5a

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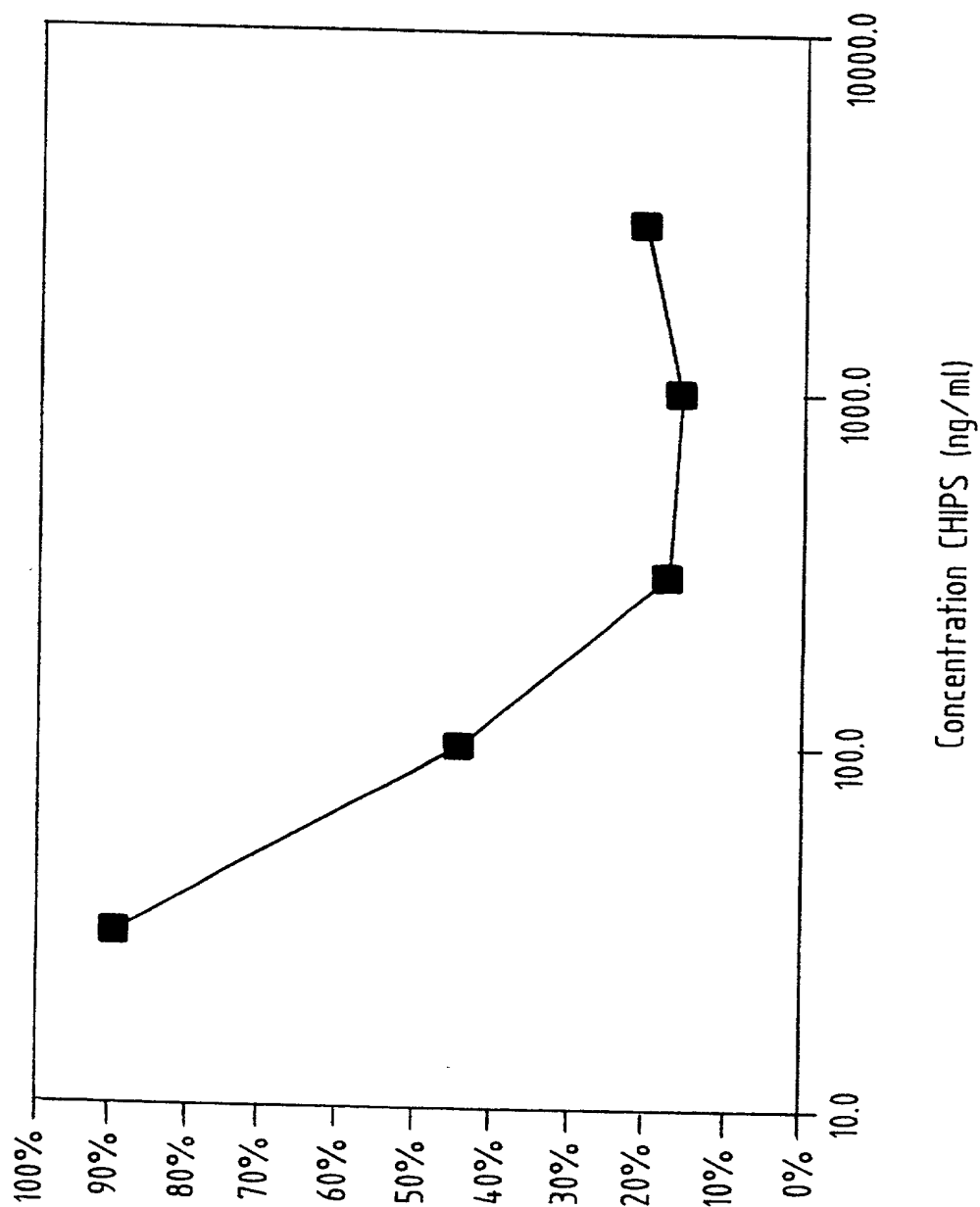
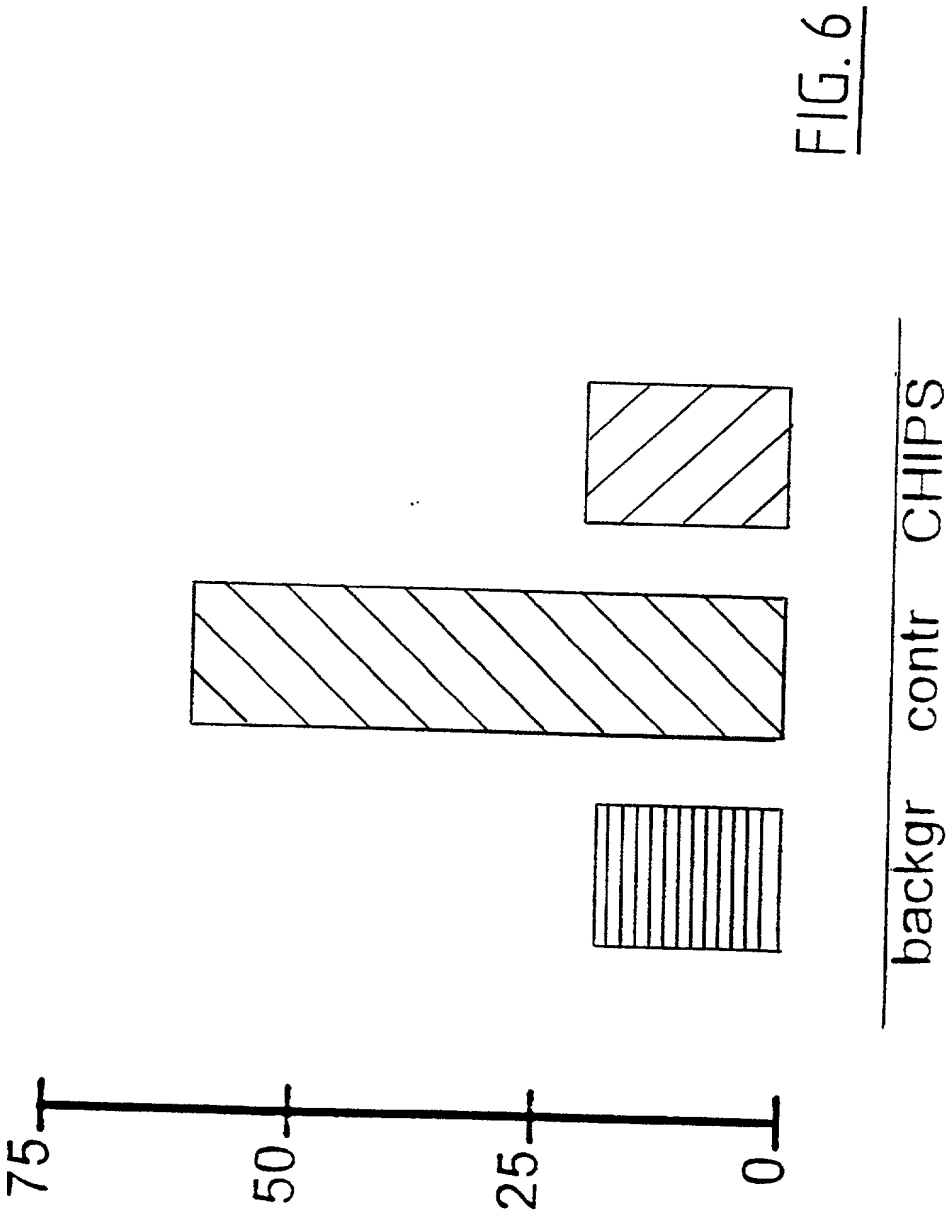


FIG. 5b



09/743364

Declaration and Power of Attorney For Patent Application

English Language Declaration

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name,

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled

Chemotaxis-inhibiting protein of staphylococcus (chips) and its use
the specification of which

(check one)

☐ is attached hereto.

☒ was filed on July 12, 1999 as PCT/NL99/00442 as
Application Serial No. 09/743,364 received on January 9, 2001
and was amended on January 9, 2001
(if applicable)

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, §1.56(a).

I hereby claim foreign priority benefits under Title 35, United States Code, §119 of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed:

Prior Foreign Application(s)

Priority Claimed

<u>1009614</u> (Number)	<u>Netherlands</u> (Country)	<u>10 July 1998</u> (Day/Month/Year Filed)
<u> </u> (Number)	<u> </u> (Country)	<u> </u> (Day/Month/Year Filed)
<u> </u> (Number)	<u> </u> (Country)	<u> </u> (Day/Month/Year Filed)

<input checked="" type="checkbox"/> Yes	<input type="checkbox"/> No
<input type="checkbox"/> Yes	<input type="checkbox"/> No
<input type="checkbox"/> Yes	<input type="checkbox"/> No

I hereby claim the benefit under Title 35, United States Code, §120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, §112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, §1.56(a) which occurred between the filing date of the prior application and the national or PCT international filing date of this application:

(Application Serial No.)

(Filing Date)

(Status)
(patented, pending, abandoned)

(Application Serial No.)

(Filing Date)

(Status)
(patented, pending, abandoned)

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith. (list name and registration number)

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